

Isolation and characterization of *in vitro* functional anti-PD-1 antibodies from ALTHEA 4:3 VH:VL Platinum Libraries™



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ABSTRACT

PD-1 (Programmed cell death protein 1) is expressed on the surface of T and B cells and has a role in down-regulating the immune system. It promotes self-tolerance by suppressing T cell inflammatory activity once coupled to its ligands PD-L1 and PD-L2, which are expressed on macrophages and dendritic cells. PD-L1 and PD-L2 are also highly expressed in several cancer cells, resulting in tumor immune evasion. Thus, the development of anti-PD1 antibodies blocking the PD-1:PD-L1 interaction has been shown to boost the immune system, promoting the destruction of cancer cells. In this work, ALTHEA 4:3 VH:VL Platinum Libraries™, a set of semi-synthetic phage display libraries, were used to generate a panel of anti-PD-1 antibodies with diverse binding, PD-1:PD-L1/PD-L2 blocking, and functional profiles. After three rounds of solution panning against recombinant PD-1, 88 clones were tested for PD-1 binding, yielding 13 positive single-chain variable fragments (scFvs), with five clones being unique. These unique clones were converted to human IgG4PE and assayed for blocking the PD-L1/PD-L2 interaction in ELISA and Jurkat cells. The antibody, D38, was selected based on its performance in these assays. The *in vitro* functionality of D38 was assessed by expression of the interferon-gamma response in a mixed lymphocyte reaction co-culture, showing a similar functional profile to FDA-approved anti-PD-1 therapeutic antibodies (Keytruda and Opdivo). These results demonstrate that ALTHEA 4:3 VH:VL Platinum Libraries™ are a valuable source for obtaining functional anti-PD1 antibodies with a promising profile for cancer drug development.

RESULTS

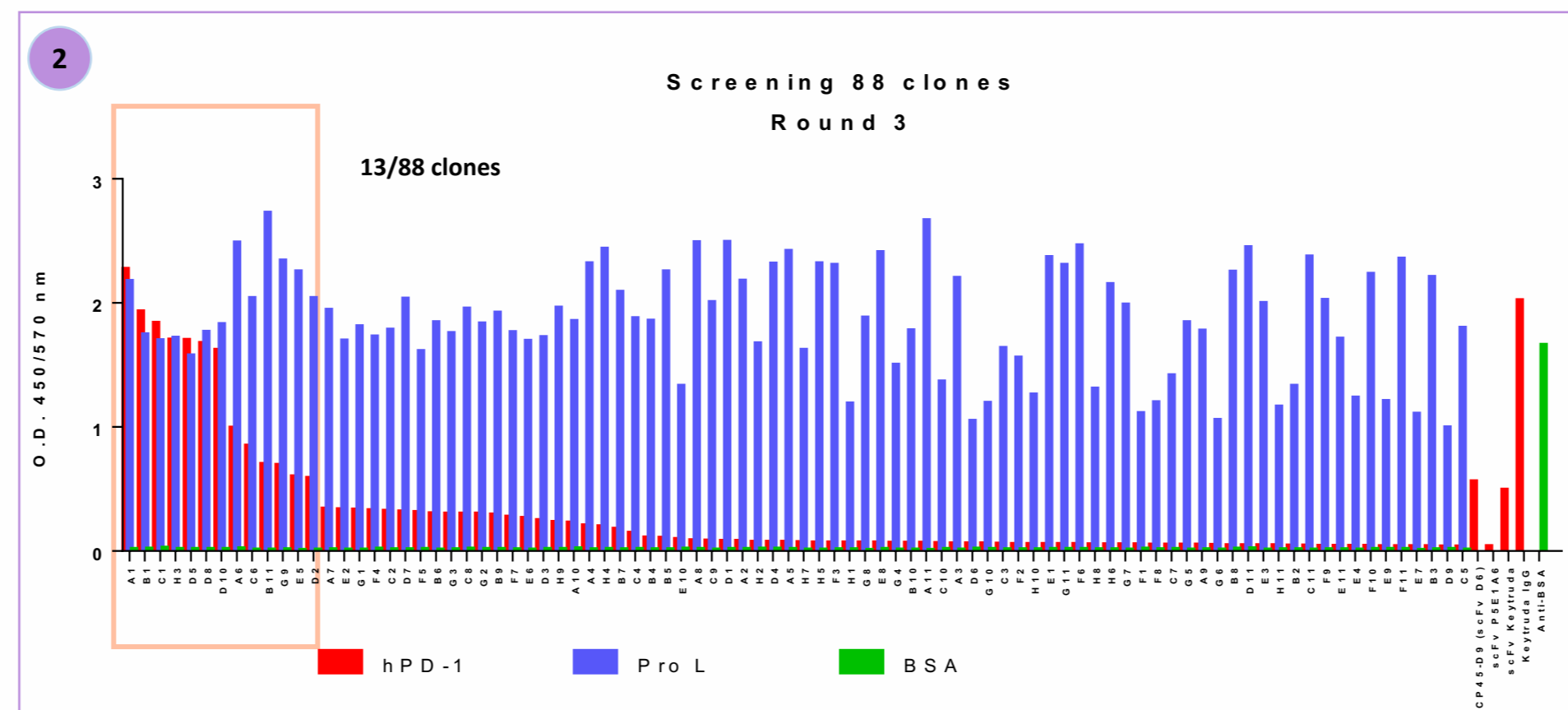


FIGURE 1. Screening 88 clones: Round 3. Evaluation of the binding to hPD-1 of 88 clones by the ELISA method.

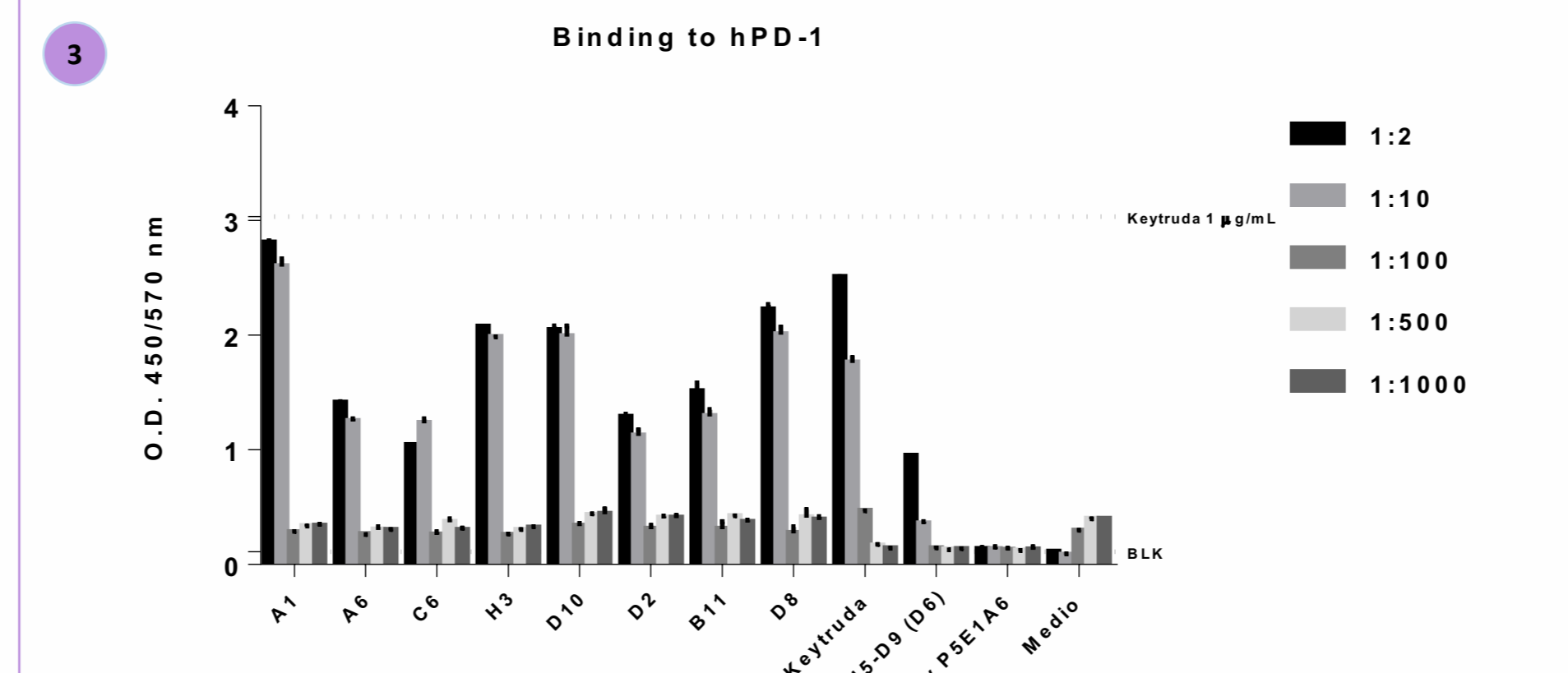


FIGURE 2. Confirmation of positive clones. Evaluation of the binding to hPD-1/ Pro L of 8 clones by the ELISA method.

IgG	Clone
D35E	PD1- IgG4 Clone A1
D36E	PD1- IgG4 Clone D2
D37E	PD1- IgG4 Clone H3
D38E	PD1- IgG4 Clone C6
D38E	PD1- IgG4 Clone B11

TABLE 1. Conversion of unique clones of Round 3 of discovery panning to IgG4.

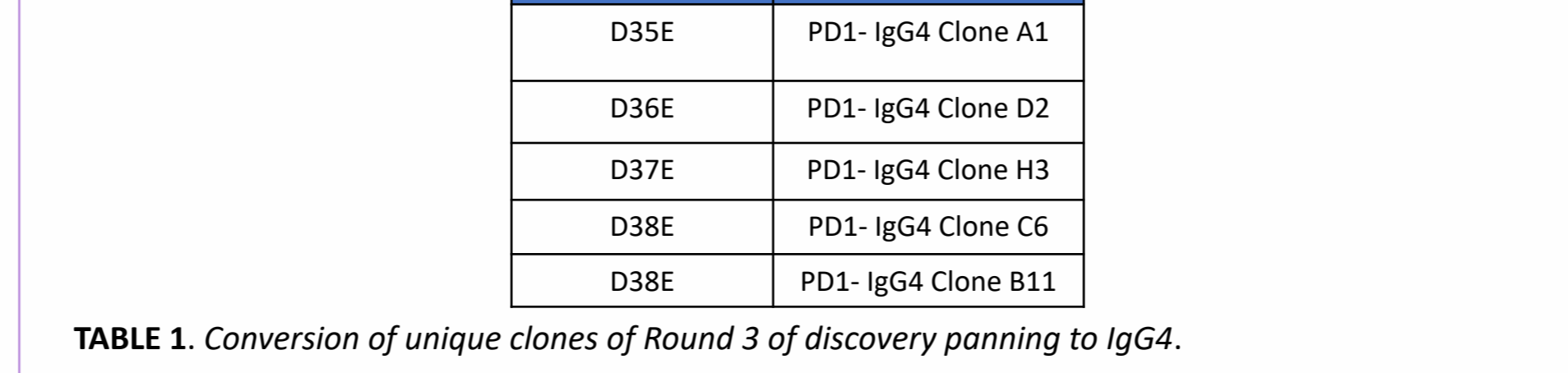


FIGURE 3. Conversion of the positive unique clones to IgG4. Evaluation of the binding of 5 clones with unique sequence to hPD-1/ Pro A by the ELISA method. For each IgG, they were transfected in duplicate in well 1 (P1) and well 2 (P2). D6E was used as a positive control (See poster No.B127).

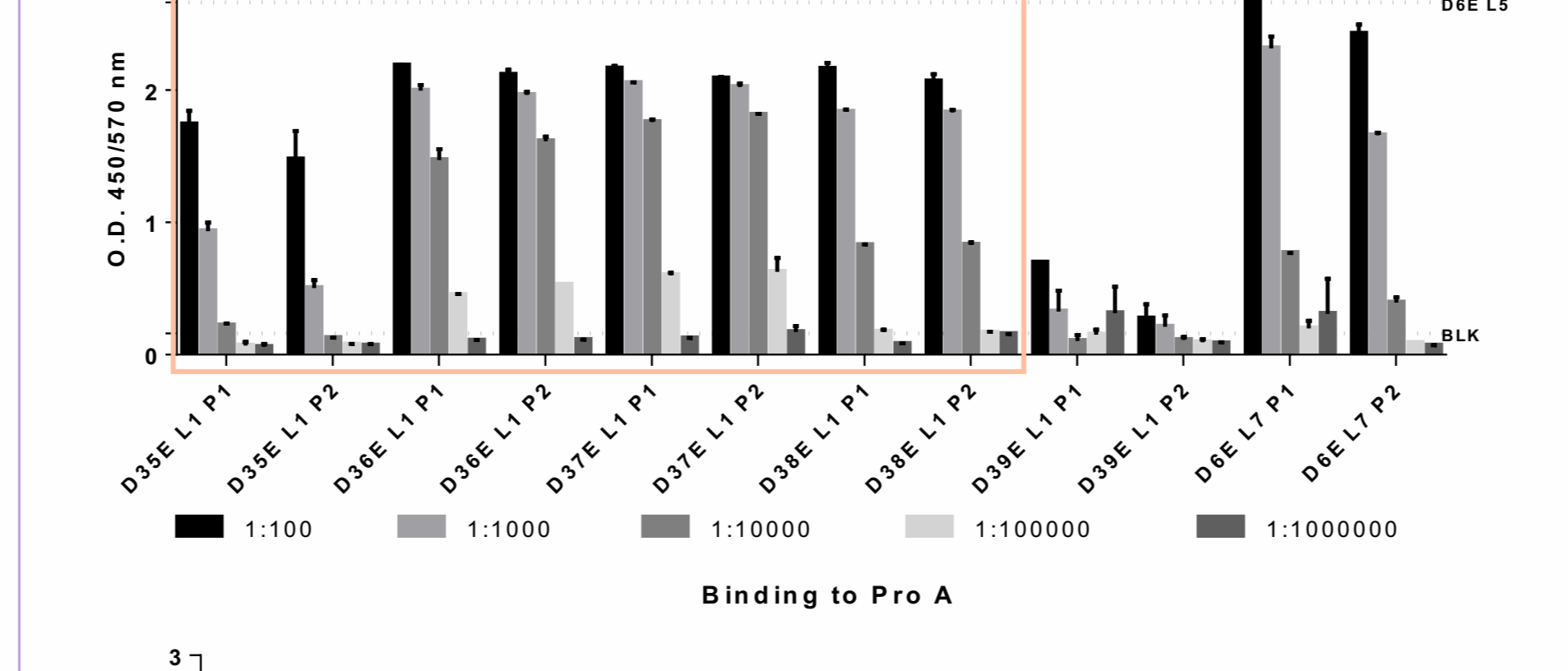


FIGURE 4. Binding to hPD-1 of purified IgG4. Evaluation of the antibodies D36E L1, D37E L1 and D38E L1 binding to hPD-1 by the ELISA method. Keytruda was used as a positive control.

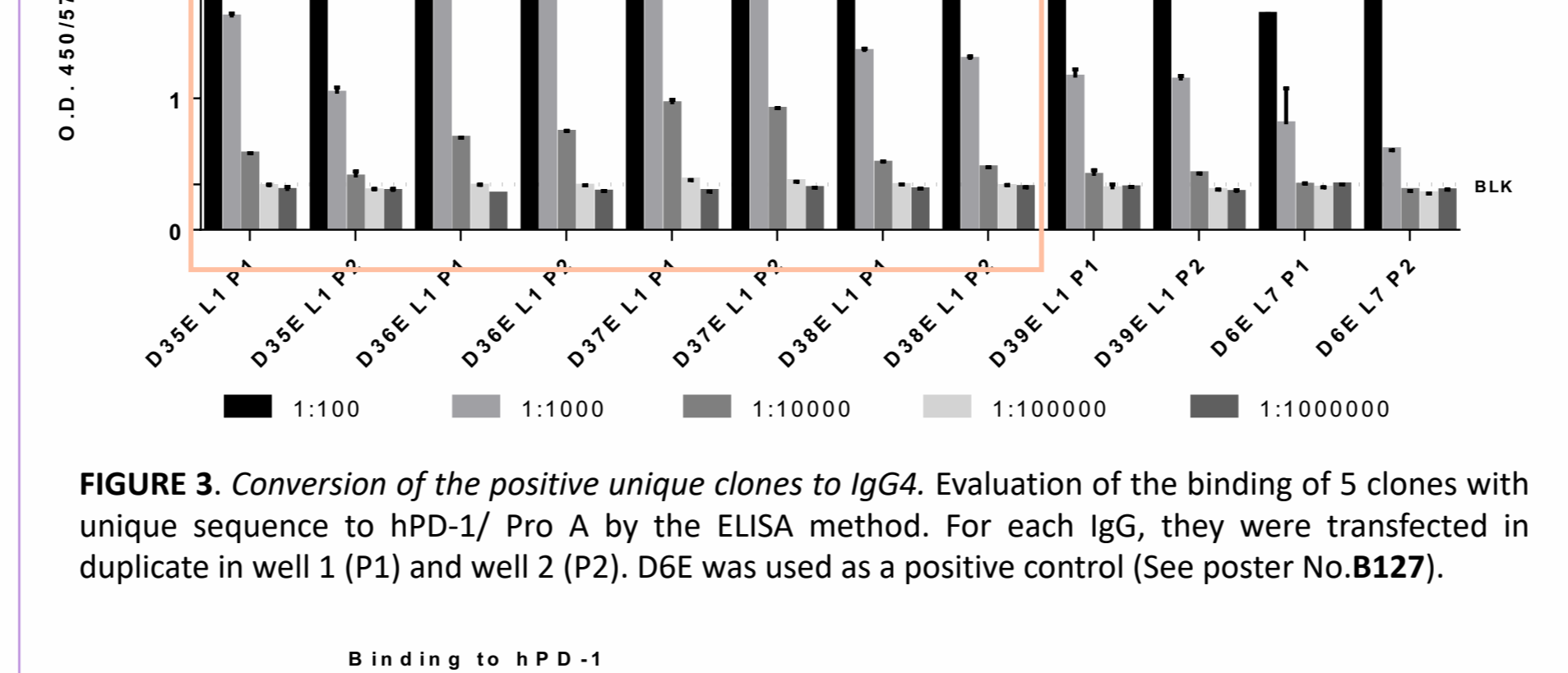


FIGURE 5. Binding to hPD-1 of purified IgG4. Evaluation of the binding of D36E L1, D37E L1 and D39E L1 to hPD-1 in Jurkat PD-1 cells. Keytruda was used as a positive control and 3C5 (anti-VEGFR3 antibody) as a negative control.

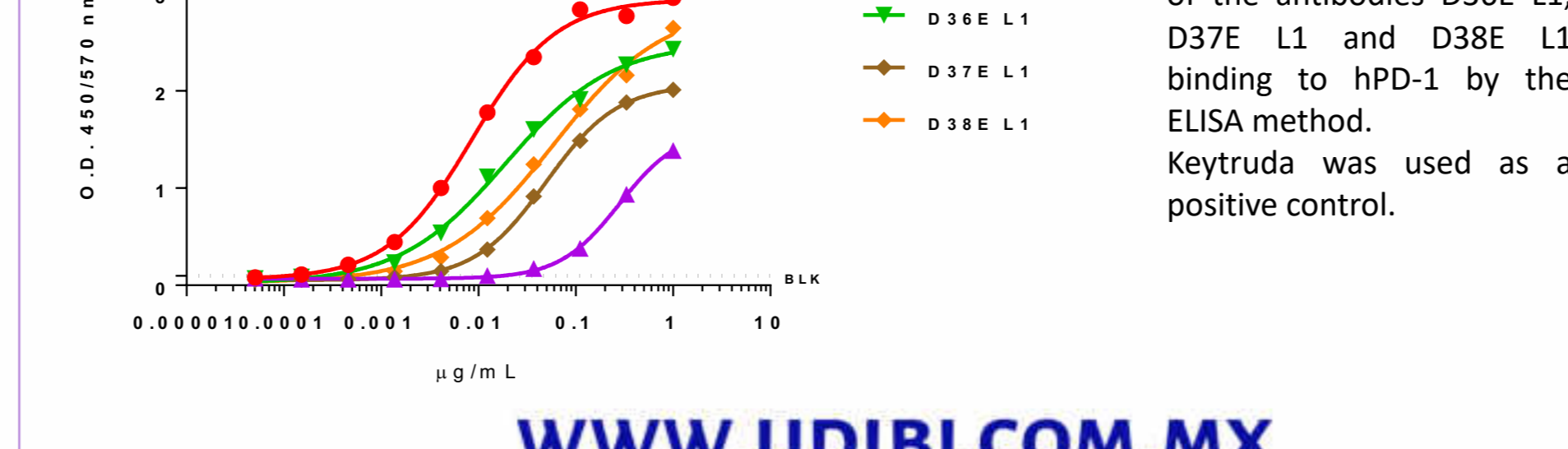


FIGURE 6. Blocking in NFAT reporter Jurkat cells. Evaluation of the blocking of D36E L1, D37E L1 and D38E L1 in NFAT reporter Jurkat cells. Keytruda was used as a positive control and 3C5 (anti-VEGFR3 antibody) as a negative control.

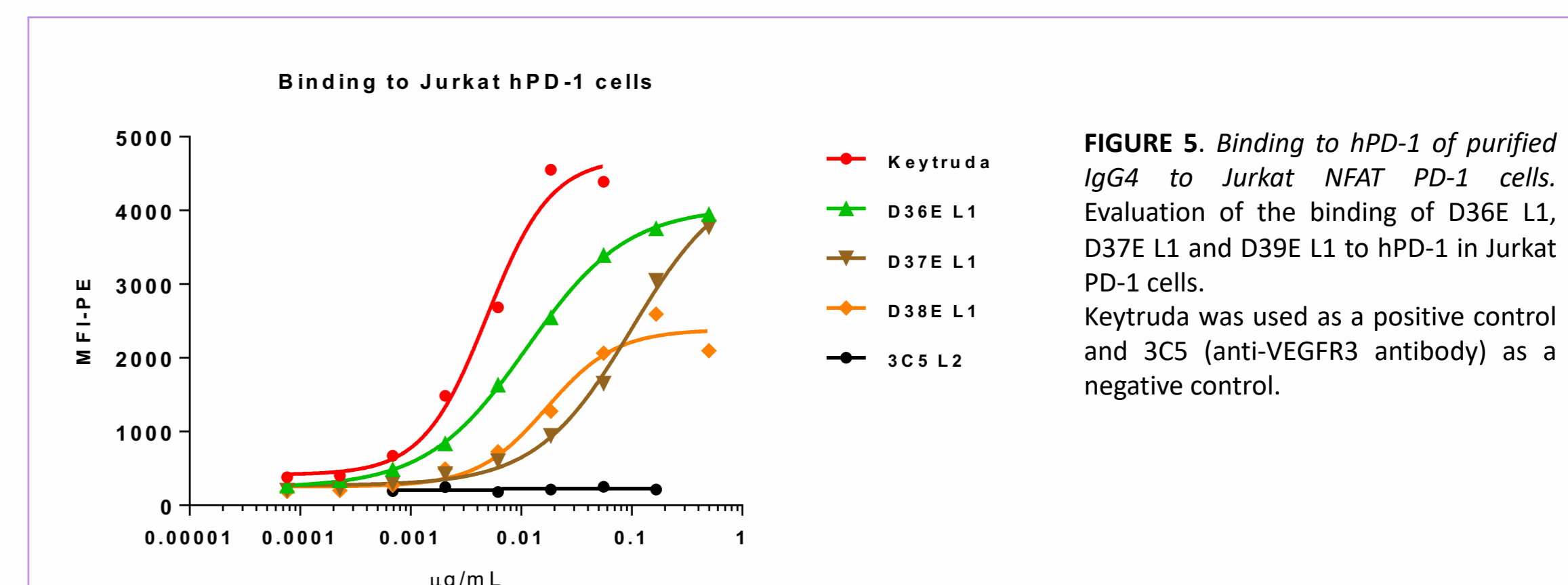
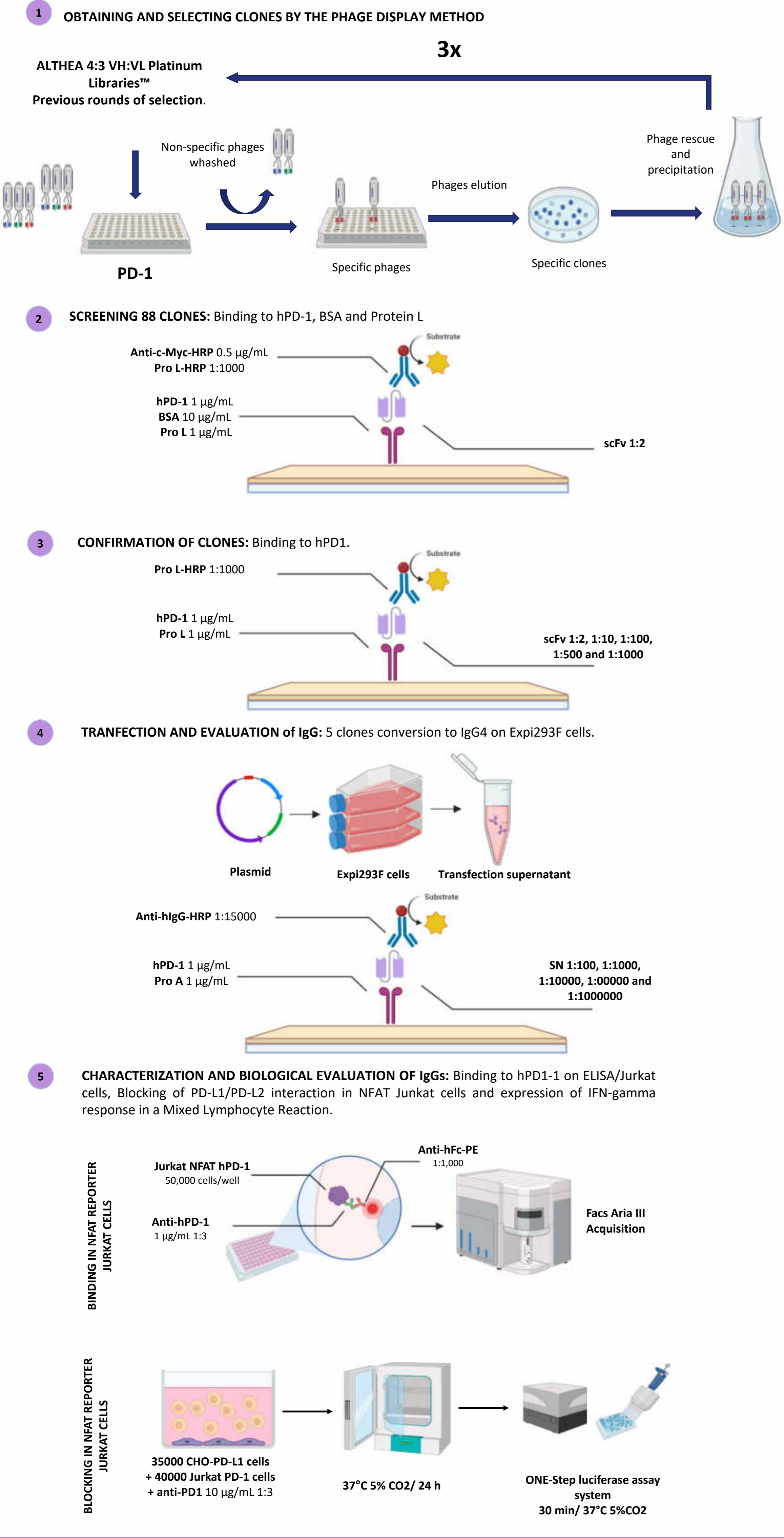


FIGURE 7. Expression of IFN-gamma response in a Mixed Lymphocyte Reaction. Evaluation of the production of IFN-gamma in a MLR. Keytruda and Nivolumab were used as positive controls and 3C5 (anti-VEGFR3 antibody) as a negative control.

METHODOLOGY



CONCLUSION

The D38 antibody obtained from ALTHEA 4:3 VH:VL Platinum Libraries™ proved to be a candidate to block the interaction *in vitro* between PD-1 and its ligand PD-L1. This activity could promote cancer cells destruction by stimulating the immune system. More importantly, it has shown to induce increased expression of cytokines such as IFN-gamma, which contributes to the antitumor immune response through its immunostimulatory and immunomodulatory effects. Thus, D38 seems to be a promising antibody to develop an anti-PD-1 therapeutic antibody to treat cancer.

REFERENCES

- Ding, G., Shen, T., Yan, C., Zhang, M., Wu, Z., & Cao, L. (2019). IFN-γ down-regulates the PD-1 expression and assist nivolumab in PD-1-blockade effect on CD8+ T-lymphocytes in pancreatic cancer. *BMC cancer*, 19(1), 1053. <https://doi.org/10.1186/s12885-019-6145-8>
- Ostrand-Rosenberg, S., Horn, L. A., & Haile, S. T. (2014). The programmed death-1 immune-suppressive pathway: barrier to antitumor immunity. *Journal of immunology (Baltimore, Md. : 1950)*, 193(8), 3835-3841. <https://doi.org/10.4049/jimmunol.1401572>
- Kwok, G., Yau, T. C., Chiu, J. W., Tse, E., & Kwong, Y. L. (2016). Pembrolizumab (Keytruda). *Human vaccines & immunotherapeutics*, 12(11), 2777-2789. <https://doi.org/10.1080/21645515.2016.1199310>
- Ghaderi, S. S., Riaz-Rad, F., Qamsari, E. S., Bagheri, S., Rahimi-Jamrani, F., & Sharifzadeh, Z. (2022). Development of a human phage display-derived anti-PD-1 scFv antibody: an attractive tool for immune checkpoint therapy. *BMC biotechnology*, 22(1), 22. <https://doi.org/10.1186/s12896-022-00752-8>
- Ostrand-Rosenberg, S., Horn, L. A., & Haile, S. T. (2014). The programmed death-1 immune-suppressive pathway: barrier to antitumor immunity. *Journal of immunology (Baltimore, Md. : 1950)*, 193(8), 3835-3841. <https://doi.org/10.4049/jimmunol.1401572>

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